

# scil v-Diro

Immunological Rapid Test

FOR VETERINARY USE ONLY!



## TEST INFORMATION

Canine heartworm disease is a chronic disease due to infection with *Dirofilaria immitis* larvae and worms. 6-7 months after infection adult female worms become apparent and can specifically be detected using the scil v-Diro assay. The clinical picture of the disease varies in affected dogs. Mild cases may be asymptomatic but more severely affected patients can show different stages of respiratory and cardiovascular diseases. The parasite is endemic in many tropical and subtropical regions. Therefore regular prophylactic treatment is needed for the dogs as well as checks for infectious status using the scil v-Diro assay.

## TEST COMPONENTS



test cassette  
(6/ 18)



Marked pipette  
(6/ 18)



reagent bottle  
(6 tests – 1x 2.5ml,  
18 tests – 1x 4.6ml)

## PLEASE NOTE PRIOR TO USE

Please use a new test cartridge for every individual test as cartridges are for single use only.

scil Rapid Test kits are for veterinary use only.

Use only test components provided by scil animal care company.

Use the test cassette within 60 minutes after opening the pouch and place the test cassette in a horizontal position on a smooth surface while the test is performed. Note the amount of sample material needed. An incorrect number of drops or too small drops may lead to false test results.

After opening the pouch, use the test cassette within one hour. Consider the test results as invalid after the read-out time.

Do not use the test after the expiration date on the pouch.

Dispose all contaminated materials properly and disinfect the work area after the test execution.

## STORAGE

scil Rapid Test kit should be stored between 2-30°C.

## REFERENCE

Kramer LH. 2009. Pathogenesis of *Dirofilaria* spp. Infections, in Proceedings of Second European *Dirofilaria* Days, 16-18 September. Morchón R, Simón F, Montoya JA, Genchi C, Eds. Salamanca, Spain: 116-123.

McCall J, et al. 2008. Heartworm and Wolbachia: therapeutic implication. *Veterinary Parasitology*, 158, 204-214.

Morchón R, Carretón E, González-Miguel J, Mellado-Hernández I. 2012. Heartworm Disease (*Dirofilaria immitis*) and Their Vectors in Europe - New Distribution Trends. *Front Physiol.*; 3():196.

Montoya-Alonso JA, et al. 2010. Canine dirofilariosis caused by *Dirofilaria immitis* is a risk factor for the human population on the island of Gran Canaria, Canary Islands, Spain. *Parasitol Res.* 107(5):1265-9.

Otranto D, et al. 2013. Vector-borne helminths of dogs and human in Europe. *Parasite & Vectors*, 6, 1-14.

## MANUFACTURER



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## SYMBOLOLOGY



Made in Germany



Expiration Date



Contents



Batch



Storage Temperature



Only for veterinary use



instruction manual

## SAMPLE MATERIAL

Best sample material is a **freshly collected serum, plasma, supernatant of whole blood or whole blood.**

Separate the **serum or plasma** from whole blood as quickly as possible. Clear, non-hemolyzed specimens can prevent a slight background staining. **Supernatant of whole blood:** Let the whole blood sample stand for some time, so that the blood sediments. The supernatant of the sedimented blood can be carefully taken up with the pipette and be used for test procedure.

A **whole blood sample** should be used as quickly as possible. **Heparin or EDTA** blood may also be used.

The sample must be at room temperature (15-25°C) and should be mixed well before used for testing.

## TEST PROCEDURE

Open the aluminium pouch, remove the test cassette. Place the test cassette on a flat surface and unscrew the bottle of reagent and place it aside.

1. Aspirate sample material with the pipette up to the mark (20 µl).

2. Apply the sample material completely on the first sample well.

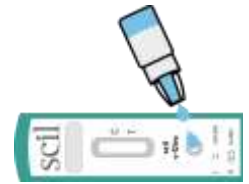
3. Add two (2) drops of the reagent from the bottle of reagent into the sample well.



Take up the sample  
with the pipette.



20 µl of sample  
in each well

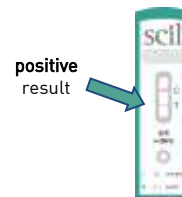


2 drops of reagent  
in each well

**Ensure that no air bubbles are formed.** If air bubbles occur, pop them with the pipette. The liquid starts running up the test strip after a short time (< 60 seconds). If whole blood is used, the fluid first needs to permeate into the test. This may take a little bit longer time than for serum. If the fluid does not run up the test strips after 90 seconds, add an additional drop of the **reagent** into the sample well, or press with the tip of the pipette into the sample well to reactivate the run of the test.

## TEST EVALUATION

The result of the test can be read after 10 minutes.



positive  
result



negative  
result



For a **positive result**, **two red lines** appear in the reaction field of the test cassette. A red line in the **T-region (T)** of the reaction field indicates a positive test result. Also a faint test line is considered as a positive test result.

The second red line in the **C-region (C)** indicates the control line, which indicates the correct performance of the test. The C-line is not a reference line and may have a different line intensity than the T-Line.

The **use of whole blood samples may lead to a lower detection sensitivity.** In case of a negative test result with whole blood, despite an existing suspicion of an infection, the test should be repeated with a serum or plasma sample from the whole blood, to obtain the maximum detection sensitivity.

## Invalid Result:

If no control line appears after the test is conducted, the test is invalid. In this case, it is likely that the test was not properly conducted or that the expiration date had already lapsed. If this occurs, a new test must be conducted.

## TEST PERFORMANCE

	Sensitivity	Specificity	Reference	n
<b>Dirofilaria</b>	96.67%**	99.99%	ELISA*	84

\*Enzyme-linked Immunosorbent Assay

\*\*compared to Knott test

